0957-4166(95)00229-4

## Total Synthesis of (S)-(+)-Curcudiol, (S)- and (R)-(-)-Curcuphenol Based on Enzymatic Resolution of a Primary Alcohol Possessing One Stereogenic Center

## Machiko Ono\*, Yuuko Ogura, Kazumi Hatogai, and Hiroyuki Akita\*

School of Pharmaceutical Science, Toho University, 2-2-1, Miyama, Funabashi, Chiba 274, Japan

**Abstract:** A highly stereoselective synthesis of the versatile chiral synthons possessing one stereogenic center, (S)-3 and (R)-3 was achieved and the application of (S)-3 and (R)-3 into the total syntheses of (S)-curcuphenol (1), (S)-curcudiol (2) and (R)-curcuphenol (1), respectively, was described.

Although numerous syntheses of racemic bisabolane sesquiterpenes have been known, <sup>1</sup> the useful asymmetric syntheses have not been reported so far except for the synthesis of (R)-(-)-curcuphenol (1) from (R)-(+)-citronellal. <sup>2</sup> Among them, (S)-(+)-curcuphenol (1), isolated from a marine sponge *Epipolasis* sp. inhibits strongly the activity of gastric H, K-ATPase, <sup>3</sup> while (R)-(-)-curcuphenol (1), isolated from a Caribbean gorgonian *Pseudopterogorgia* rigida and *Lasianthaea podocephala* exhabits antibacterial activities against *Staphylococcus aureus* and *Vibrio anguillarum*. <sup>4</sup> We now report that (S)-1, (R)-1, and (S)-curcudiol (2) have been synthesised based on enzymatic resolution using immobilized lipase in organic solvent.

The most intriguing point of the present synthesis is the preparation of the optically active primary alcohols possessing one stereogenic center eq. (S)- and (R)-3. This was successfully achieved by carrying out an enantioselective hydrolysis of  $(\pm)$ -acetate 4 obtained by the acetylation of  $(\pm)$ -3, using immobilized lipase. The desired racemic  $(\pm)$ -3 was already obtained in the reaction of methyl (4,5)-epoxy-(2E)-pentenoate and m-methoxytoluene in the presence of BF3.Et2O by us.<sup>5</sup>

Me

OMe

OMe

$$BF_3 \cdot Et_2O$$

OMe

RO

COOMe

 $R = H$  (±)-3

 $R = Ac$  (±)-4

1830 M. Ono et al.

Table

Initially, (±)-4 was subjected to screening experiments using several kinds of commercially available lipases. Among them, two lipases "MY-30" from Candida cylindracea and "OF-360" from Candida cylindracea were found to be effective. When (±)-4 was subjected to the enantioselective hydrolysis using "MY-30" in water saturated isopropyl ether, an alcohol (S)-3 (27%, 80% ee)<sup>6</sup> and the unchanged (R)-4 (69%, 36% ee) were obtained. On the other hand, asymmetric hydrolysis of (±)-4 using "OF-360" gave (S)-3 (60%, 51% ee) and (R)-4 (38%, 83% ee). The desired stereochemistry of 3 was found to be governed by the selection of lipase. Then immobilized lipases "MY-30" and "OF-360" were obtained by illumination of a mixture consisting of a photo-crosslinkable resin prepolymer ENTP-4000, 7 a photo-sensitizer such as benzoin ethyl ether and the crude lipases "MY-30" and "OF-360", respectively. Using the immobilized lipases afforded the much better results as shown in table [entry 2, (S)-3, 85% ee; entry 4, (R)-4, 90% cel. The alcohol (S)-3 having 80% enantiomeric excess was subjected to the enantioselective acetylation using "OF-360" in the presence of isopropenyl acetate in isopropyl ether to afford (S)-4 (74%, 90% ec,  $[\alpha]p$  -7.2, c=0.46, McOH) and (R)-3 (16%, 30% ee). Treatment of (S)-4 and (R)-4 (entry 4, 90% ee) with MeONa in MeOH produced (S)-3 (90% ee, [a]p -15.1, c=0.67, MeOH) and (R)-3 (90% ee), respectively. The enantiomeric purity of the obtained chiral compounds was determined by HPLC on a CHIRALCEL OD (250 X 4.6 mm) column. In order to confirm the absolute configuration of the present (-)-3, (-)-3 was successfully converted to the reported acid (S)-5.8 Thus the absolute structure of (-)-3 was determined to be S. Then total synthese of (S)-curcudiol 2, (R)-curcuphenol (1) and (S)-1 formally derived from (S)-2, were achieved from (S)-3 (90% ee) and (R)-3 (90% ee), respectively. Catalytic hydrogenation of (S)-3 gave (S)-6 followed by treatment of toxyl chloride to afford (S)-7 [83% overall yield from (S)-3]. NaBH4 reduction of (S)-7 provided (S)-8 (42%) and (S)-9 (40%). Conversion of (S)-9 into the one-carbon homologation product (S)-11 was achieved by the standard procedure [iodination, (S)-10

		COOMe RO	Me OM	+	Me OMe COOMe	
R=Ac (±)-4 $R=Ac$ (F			R=Ac (R)-4	R=H (S)-3		
R=H (S)-3			R=H (R)-3		R=Ac (S)-4	
Entr	y Substrate(g)	Lipase		Prod %(% ee)		
1	(±)-4 (0.2)	MY-30 (Candida cylinda	racea)	(R)-4 69(36)	(S)-3 27(80)	
2	(±)-4 (0.25)	Immobilized lipase (MY	-30)	(R)-4 77(24)	(S)-3 22(85)	
3	(±)-4 (0.2)	OF-360 (Candida cylind	iracea)	(R)-4 38(83)	(S)-3 60(51)	
4	(±)-4 (0.2)	immobilized lipase (OF-	-360)	(R)-4 40(90)	(S)-3 52(58)	
5*	(S)-3 (0.1)	OF-360		(S)-4 74(90)	(R)-3 16(30)	

<sup>\*</sup> Optically active (S)-3 (80% ee) was employed.

a; H<sub>2</sub>/Pd(OH)<sub>2</sub>-C b; TsCl/pyridine c; NaBH<sub>4</sub>/DMSO d; l<sub>2</sub>/PPh<sub>3</sub>

e; NaCN/DMF f; 1) NaOH/EtOH 2) CH<sub>2</sub>N<sub>2</sub> g; EtSH/AICI<sub>3</sub> h; MeMgI/Et<sub>2</sub>O

i; Ac<sub>2</sub>O/pyridine j; MeOCH<sub>2</sub>Cl/NaH/18-Crown-6 k; 1N-NaOH/MeOH

I; PCC/CH<sub>2</sub>Cl<sub>2</sub> m; Ph<sub>3</sub>P<sup>+</sup>-CHMe<sub>2</sub> I /n-BuLi n; 2N-HCI/i-PrOH

1832 M. Ono et al.

(48%), CN-substitution, (S)-11 (99%)]. An alkaline hydrolysis of (S)-11 followed by the successive esterification gave the methyl ester (S)-12 (54% overall yield from (S)-11,  $[\alpha]_D$  +6.2, c=1.15, MeOH). Demethylation of (S)-12 with a combination of AlCl3 and EtSH provided a phenol (S)-13, which was treated with Grignard reagent to afford (S)-curcudiol (2) ([\alpha]D +9.9, c=4.96, CHCl3; corresponds to 90\% ee) in 91\% overall yield from (S)-12. The spectral data ( $[\alpha]_D$ ,  ${}^1H$ -NMR and  ${}^{13}C$ -NMR) of the synthesized (S)-2 were identical with those (\( \begin{align\*} \lambda \rightarrow \partial \text{p} + 9.2, c=10.8, CHC\( \alpha \rightarrow \rightarrow \text{of natural (S)-2, which is converted into (S)-curcuphenol (S) and (S) are the converted into (S) are the converted into (S) and (S) are the converted into (1) in the literature.<sup>3</sup> The synthesis of (R)-curcuphenol (1) from (R)-3 (90% ee) was carried out foundamentally by the same way as that of (S)-3. Conversion of (R)-3 into the alcohol (R)-9 was achieved by the same route [(R)-7, 83% overall yield from (R)-3, (R)-8 (47%) and (R)-9 (26%)] as the previous case. Acetylation [(R)-14, 90%] of (R)-9 followed by demethylation provided the phenol (R)-15 (97%), which was treated with methoxymethyl chloride (MOMCl) to give the MOM ether (R)-16 in 97% yield. Hydrolysis [(R)-17, 98%] of (R)-16 followed by oxidation provided the aldehyde [(R)-18], which was subjected to the Wittig reaction to afford (R)-19 [62% overall yield from (R)-17]]. Deprotection of (R)-19 gave (R)-curcuphenol (1) (62% yield, [α]p -20.9, c=1.73, CHCl3; corresponds to 90% ee), whose spectral data ([α]p, ¹H-NMR and <sup>13</sup>C-NMR) were identical with those ( $[\alpha]_D$  -23.6, CHCl<sub>3</sub>) <sup>2</sup> of natural product (R)-1.

Acknowledgement: The authors are grateful to Professor N. Fusetani, Tokyo University, Japan for generously providing the spectral data of natural (S)-(+)-curcuphenol (1). This work was supported by a grant for the Biodesign Research Program from the Institute of Physical and Chemical Research (RIKEN) to H. A. and a Grant-in-Aid for Scientific Research (No. 06672115) from the Ministry of Education, Science and Culture of Japan to H. A.

## References and Notes

- 1) J. ApSimon, "The Total Synthesis of Natural Products," vol 5, John Wiley and Sons, 1983, p 35.
- 2) E. L. Ghisalberti, P. R. Jefferies and A. D. Stuart, Aust. J. Chem., 32, 1627 (1979).
- 3) a) N. Fusctani, M. Sugano, S. Matsunaga and K. Hashimoto, Experientia, 43, 1234 (1987).
  - b) A. E. Wright, S. A. Pomponi, O. J. McConnell, S. Kohmoto and P. J. McCarthy, J. Nat. Prod., 50, 976 (1987).
- 4) F. J. McEnroe and W. Fenical, Tetrahedron, 34, 1661 (1978).
- 5) M. Ono, R. Todoriki, Y. Yamamoto and H. Akita, Chem. Pharm. Bull., 42, 1590 (1994).
- 6) Satisfactory analytical data were obtained for all new compounds.
- 7) S. Fukui and A. Tanaka, Advances in Biochemical Engineering/Biotechnology, 29, 1 (1984).
- 8) L. Verbit, A. S. Rao and J. W. Clark-Lewis, *Tetrahedron*, **24**, 5839 (1968). A detailed conversion procedure will be reported in the forthcoming paper.